

# Methicillin-resistant *Staphylococcus aureus* infective endocarditis

**Pascal M. Dohmen**

Dept. of Cardiac Surgery, Heart Centre Leipzig, University of Leipzig, D-04289 Leipzig, Germany.

Address reprint requests Pascal M. Dohmen MD PhD, Department of Cardiac Surgery, Heart Centre Leipzig, University of Leipzig, Struempellstrasse 39, D-04289 Leipzig, Germany. Telephone +49 341 865 1422 Fax +49 341 865 1452 E-mail: [pascal.dohmen@yahoo.de](mailto:pascal.dohmen@yahoo.de)

Methicillin was introduced in 1959 to treat penicillin-resistant *S. aureus* infection, however Methicillin-resistant *S. aureus* (MRSA) was reported 2 years later (1). The most common organism identified in acute infective endocarditis (AIE) is still *S. aureus* associated with increased hospital mortality, morbidity and hospitalization. In Germany the proportion of MRSA among *S. aureus* microbiologic examinations was 1.1% in 1990 and dramatically increased to 17.5% by 2001 and reached 21.9% in 2009 (2). Due to the biofilm in AIE, adequate treatment is challenging, including appropriate antibiotic therapy. To improve outcome of AIE early adequate antibiotic therapy is needed, however vancomycin has a decrease microbe clearance and poor clinical response compared with  $\beta$ -lactam agents in Methicillin-susceptible *S. aureus*. Today, several reports on clinical failures in MRSA with vancomycin were documented even at MICs <2.0  $\mu$ g/ml. Current guidelines recommend higher trough vancomycin of 10-20  $\mu$ g/ml, which will lead to increase complications such as nephrotoxicity (3). Therefore more effective alternative antibiotic treatment is needed to treat MRSA endocarditis and reach sufficient into the biofilm (4). This overview reports recent developments to improve MRSA endocarditis treatment.

**Keywords** MRSA, active infective endocarditis, biofilm

## 1. Introduction

Endocarditis was first described in 1885 by Sir William Osler (5), which is a demanding disease in which a high number of patients require surgery due to antibiotic therapy failure (6). Generally active infective endocarditis (AIE) has a high mortality rate and therefore many studies have been performed to identification risk factors, clinical improvements, predictors, including the modified Duke criteria which should guide and improve the treatment of AIE (7). Nevertheless in-hospital mortality has remained high and has not varied substantially during the last decades. Additionally patients risk profiles has increased during the last years and contribute to a continuing high mortality rate.

One important factor in AIE is the caused microorganism of which *Staphylococcus aureus* (*S. aureus*) show a poorer outcome compared to other patients suffering from different microbes. In fact it is an independent risk factor for increased morbidity and mortality (8). Data from the International Collaboration on Endocarditis showed that in-hospital mortality, which occurred in 22.8% of the study patients, was predicted by older age, health-care-associated infection, *S. aureus* infection and complications of PVE, including heart failure, stroke, intra-cardiac abscess and persistent bacteraemia (9). The authors concluded also that complications of PVE strongly predict in-hospital mortality, which remains high despite prompt diagnosis and the frequent use of surgical intervention (10).

*S. aureus* shows not only higher morbidity and mortality rates, but is also the most common cause of AIE (11,12). The increased complication associated with *S. aureus* is due to the highly invasive character with early tissue infiltration, extending to excessive abscess formation (13). Furthermore Corral et al (14) found that *S. aureus* in native valve endocarditis statistically significantly increased the risk of neurological complications (p=0.024).

In 1959, methicillin was introduced to treat penicillin-resistant *S. aureus* infections, however after two years first methicillin-resistant *S. aureus* (MRSA) were found (1). In Germany the proportion of MRSA among *S. aureus* microbiologic examinations was 1.1% in 1990 and dramatically increased to 17.5% by 2001 and reached 21.9% in 2009 (2). To improve outcome of AIE early adequate antibiotic therapy is needed, however vancomycin which is used in MRSA endocarditis has a decrease microbe clearance and poor clinical response compared with  $\beta$ -lactam agents in methicillin-susceptible *S. aureus* (MSSA). Therefore vancomycin should be avoided in patients with MSSA and can only be used after methicillin-resistance has been proven. Furthermore *S. aureus* within the biofilms have several defense mechanisms against antimicrobial agents (15). Therefore several alternative antibiotic therapies have been introduced and forward in guidelines to treat AIE today (16,17). An extreme demanding problem is still the treatment of MRSA endocarditis, which is a combination of surgery and supported by a limited number of antimicrobial agents. To treat AIE adequately it is important to understand the complexity of this disease, starting with the biofilm.

## 2. The Biofilm

Biofilms have been described as early as 1684 by Anton van Leeuwenhoek, the so-called "animalculi", in the plaque on his own teeth investigated with his developed microscope. Nevertheless it took almost three hundred years until the theory of biofilm was promulgated (18). The biofilm is defined as "an aggregate of microbial cells adherent to a living

or non-living surface, embedded within a matrix of extracellular polymeric substances of microbial origin" (19). Biofilms are more resistant to antibiotic therapy and are protected by different mechanism such as reduced penetration of antibiotics, protective surface, bacteria within the biofilm are slow growing with reduced DNA synthetic activity, showing stress response genes and antigen expression, increased mutation rates, persister cells,  $\beta$ -lactamase receptors control and decrease infiltration due to be a diffusion barrier (15). *S. aureus* are dominant cause of biofilm-associated infections in which *S. aureus* involves most often host tissues (20), which produce a multi-layered biofilm, including glycocalyx, a specific polysaccharide antigens called polysaccharide intercellular antigen (PIA). Important is the different regulation of Biofilms development in MSSA and MRSA. O'Neill and colleagues (21) found that MRSA biofilms are more regulated by SarA and Agr adhesin proteins and *ica* independent, whereas MSSA biofilms development is linked by SarA-regulated PIA and polymeric N-acetyl-glucosamine PNAG.

Jefferson et al (22) studied the reduced penetration of glycopeptide antibiotic e.g. vancomycin into viable *S. aureus* within a biofilm using confocal scanning laser microscopy with fluorescently labeled derivative of vancomycin. This study suggests that, whereas planktonic bacteria were rapidly exposed to a full bolus of vancomycin, the bacteria in the deeper layers were exposed to a gradually increasing dose of the drug. Therefore bacteria living within a biofilm undergo stress induced metabolic or transcriptional changes that increase resistance to the antibiotic. PNAG, an important component of the *S. aureus* biofilm matrix, was not involved in the observed decrease in the rate of vancomycin penetration.

Rani et al (23) investigated different patterns of DNA replication and protein synthetic activity of staphylococcal biofilms by using inducible green fluorescent protein construct. His study suggested that staphylococcal biofilms contain cells in at least four distinct states: growing aerobically, growing fermentation, dead, and dormant. The variety of activity states represented in a biofilm may contribute to the special ecology and tolerance to antimicrobial agents of biofilms.

Bagge et al (24) showed that patients suffer from cystic lung fibrosis, who are generally colonized with *Pseudomonas aeruginosa* biofilms, show an effect on subinhibitory concentrations of a  $\beta$ -lactam antibiotic, such as imipenem. In total 34 genes showed statistically significant differential expression in response to imipenem, induced or repressed in biofilms exposed to imipenem compared to the controls. The strong impact was induced by *ampC*, which codes for chromosomal  $\beta$ -lactamase.

Oliver A et al (25) investigated chronic infections in patients with cystic lung fibrosis caused by *P. aeruginosa*, having increased adaptation on antibiotic therapy. This was explained by spontaneous mutation rates, in this study found by 36% with hypermutable (mutator) strains and the evolution of antibiotic resistance.

Lechner et al (26) studied bacterial persister cells, which are non- or slow-growing reversible phenotypic variants of the wild type, showing an increased tolerance to bactericidal antibiotics. This study investigated *S. aureus* persister levels by monitoring colony-forming unit counts of planktonically grown cells treated with different antimicrobials over time. The study showed that *S. aureus* in the stationary phase are equivalent to persister cells, as not all of these cells showed antibiotic tolerance.

Bagge et al (27) studied the expression of chromosomal AmpC  $\beta$ -lactamase in *P. aeruginosa* regulation by the activity of an amidase, AmpD. Some *P. aeruginosa* strains show resistant variants isolated from in vivo and in vitro biofilms for mutations in ampD leading to high-level expression of chromosomal  $\beta$ -lactamase. Identical mutation in *Enterobacter cloacae* were found of AmpR which cause a 450-fold higher AmpC expression. This pathway can also lead to an increase antimicrobial resistance in biofilms.

Xu et al (28) investigated the effect of decrease infiltration due to be a diffusion barrier for antimicrobial agents. De Beer et al (29) evaluated the penetration of chlorine into biofilms during an ongoing infection. This study showed the deactivation of antimicrobial agents in the upper surface layers of the biofilm before the deeper layer were reached. A recent study of Singh et al (30) evaluated the penetration of different antibiotics such as oxacillin, cefotaxime and vancomycin in *S. aureus* and *S. epidermidis* biofilms, finding similar results.

### 3. Antibiotic therapy options

The different antimicrobial agents to treat MRSA endocarditis are limited, however the success of the treatment is not only depending on selecting the correct substance but timing of an appropriate antibiotic therapy is also essential. The Tarragona strategy says "hit hard and early" (31), which will have an enormous influence on the mortality. Schramm et al (32) found in a multi-variant analysis that inappropriate antimicrobial treatment is one independent risk factor for hospital mortality (adjusted Odds Ratio (OR) 1.92 with 95% confidence interval (CI) 1.48-2.50; p-value 0.013). Cheong HS et al (33) showed that inappropriate antibiotic therapy will have a negative influence on the survival curve at the Kaplan-Meier analyses (p=0.050). Furthermore the working mechanism of an antimicrobial agent is important, showing a bacteriostatic or bactericidal effect (34). Pankey GA et al (35) demonstrated the advantages of bacteriostatic over bactericidal activity who will avoid toxic shock syndrome due to exotoxins or part of the microbial components. On the other hand their activity is not active and limited against microbes and less effective in biofilm, which are available in AIE.

#### 4. Vancomycin

Vancomycin is the golden standard in MRSA AIE treatment, generally used in combination with other antimicrobial agents. Although this agent is widely applied, it only limitedly penetrates tissue or biofilm, has slow bactericidal activity, and has adverse effects such as Red man syndrome, neutropenia, thrombocytopenia and renal failure (36). Soriano et al (37) studied the influence of minimal inhibitory concentration (MIC) on MRSA bacteremia. In a logistic regression model this study showed that MICs of  $> 2 \mu\text{g/mL}$  had a significant higher failure rate (OR, 3.62; 95% CI 1.20-10.9). Lodise and colleagues (38) compared the outcome between low and high vancomycin MICs in bacteremia with MIC  $\geq 1.5 \mu\text{g/mL}$  (n=66) and MIC  $< 1.5 \mu\text{g/mL}$  (n=26) and an overall failure rate of respectively 36.4% versus 15.4% (p=0.49) and hospitalization duration stay of respectively median 21 (range 9-43) and 11 (range 9-17) (p=0.02). Switch to alternative antibiotic therapy was in 20% and 2% respectively needed. This results showed a trend, however was not statistically significant. Vancomycin resistant *S. aureus* is very rare, however the number of patients vancomycin-intermediate (VISA) and heterogeneous VISA are growing (39,40).

#### 5. Teicoplanin

Teicoplanin is a semisynthetic glycopeptide, similar to vancomycin, however has no renal toxicity. There are studies which show equal results with teicoplanin and vancomycin, however teicoplanin is generally better tolerated. Rolston et al (41) performed a prospective, randomized, double-blind study comparing teicoplanin and vancomycin for the treatment of gram-positive bacteremias in neutropenic patients. This study also showed more adverse reactions occurred more often in the vancomycin group (31%) than in the teicoplanin group (9%; P = .06). Huang et al (42) performed so far the only retrospective study using teicoplanin versus vancomycin in MRSA endocarditis, however could not find any statistically significant difference in hospital mortality rate and therapy failure.

#### 6. Tigecycline

Tigecycline is a glycylcycline antimicrobial agent with bacteriostatic effect against *S. aureus* including MRSA. This antibiotic has limited side-effects, however mainly vomiting and nausea which can be dose limited. Due to the peak serum concentration it is only limitedly effective in bacteremia (36). On MRSA endocarditis there are no studies available.

#### 7. Linezolid

Linezolid is an oxazolidinone is bacteriostatic against staphylococci. The advantage of this antimicrobial agent is the availability for oral and intravenous application. There are some disadvantages of linezolid as it is limited to 28 days of application. Bone marrow suppression, thrombocytopenia and irreversible sensory and motor polyneuropathy have been reported as side-effects (36). Wilcox et al (43) showed in a Phase 3 study using linezolid in catheter-associated bacteremia a higher mortality than with vancomycin.

#### 8. Telavancin

Telavancin is a lipoglycopeptide which needs to be applied once a day. It is effective against MRSA, however also against VISA and VRSA. No clinical studies are available on MRSA endocarditis, however Smith et al showed some superiority in a range of in vitro biofilms models of multi-resistant *S. aureus* (44).

#### 9. Daptomycin

Daptomycin is a cyclic lipopeptide with rapid bactericidal effect against *S. aureus* including MRSA. Although it is bactericidal it does not release endotoxin which will lead to toxic shock syndrome. The side effect of daptomycin is rhabdomyolysis. Fowler et al (45) showed a similar effect on complicated bacteremia from daptomycin versus combined standard therapy, however there was a significant decrease on renal complications. A recent study of Dohmen et al (4) showed favourable results of daptomycin in Endocarditis including a high number of patients with MRSA endocarditis. Today there are several studies on daptomycin use in resistant and multi-resistant endocarditis which has also been included in several guidelines (16,17).

## 10. Surgical options

Current guidelines suggest that if surgery is performed short-term mortality will be decrease, in patients with specific symptoms or organisms; however it is unclear when surgery should be performed (46). Head et al (46) performed a meta-analysis on the current data available on medical or surgical therapy in AIE. These data support that surgery can decrease in-hospital mortality with an overall odds ratio of 0.47 (CI 0.38-0.58). however a marked statistically significant heterogeneity was found ( $I^2 = 65\%$ ,  $p = 0.005$ ) which means there is an excessive variation of compared results.

The first clinical trial, so called ENDOVAL is a randomized prospective multi-centre study to compare medical and surgery treatment in patients suffering from AIE. (47)

AIE caused by *S. aureus* are challenging and complicated due to large vegetations and embolic manifestations with increased mortality and if a multi-resistant *S. aureus* is found, surgery will always be indicated (46).

## References

- [1] Jevons MP. Celbenin-resistant staphylococci. *BMJ* 1961;124-6.
- [2] Koeck R, Mellmann A, Schaumberg F, Friedrich AW, Kipp F, Becker K. The epidemiology of methicillin-resistant staphylococcus aureus (MRSA) in Germany. *Dtsch Arztebl Int* 2011;108:761-7.
- [3] Borde JP, Kern WV. Therapie von MRSA-Infektionen. *Dtsch Med Wochenschr* 2012;137:2553-7.
- [4] Dohmen PM, Guleri A, Capone A, Utili R, Seaton RA, Gonzalez-Ramallo VJ, Pathan R, Heep M, Chaves RL. Daptomycin for the treatment of infective endocarditis: results from a European registry. *J Antimicrob Chemother* 2013;68:936-42.
- [5] Osler W. The Gulstonian Lectures, on Malignant Endocarditis. *Br Med J* 1885;1:467-70.
- [6] Tornos P, Jung B, Permanyer-Miralda G, Baron G, Delahaye F, Gohlke-Barwolf C, Butchart EG, Ravaud P, Vahanian A. Infective endocarditis in Europe: lessons from the Euro heart survey. *Heart* 2005;91:571-5.
- [7] Durack DT, Lukes AS, Bright DK. New criteria for diagnosis of infective endocarditis: utilization of specific echocardiographic findings. Duke Endocarditis Service. *Am J Med.* 1994;96:200-9.
- [8] Nomura A, Omata F, Furukawa K. Risk factors of mid-term mortality of patients with infective endocarditis. *Eur J Clin Microbiol Infect Dis* 2010;29:1355-60.
- [9] Durante-Mangoni E, Bradley S, Selton-Suty C, Tripodi MF, Barsic B, Bouza E, Cabell CH, Ramos AI, Fowler V Jr, Hoen B, Konečný P, Moreno A, Murdoch D, Pappas P, Sexton DJ, Spelman D, Tattevin P, Miró JM, van der Meer JT, Utili R; International Collaboration on Endocarditis Prospective Cohort Study Group. Current features of infective endocarditis in elderly patients: results of the International Collaboration on Endocarditis Prospective Cohort Study. *Arch Intern Med.* 2008;168:2095-103.
- [10] San Román JA, López J, Vilacosta I, Luaces M, Sarriá C, Revilla A, et al. Prognostic stratification of patients with left-sided endocarditis determined at admission. *Am J Med.* 2007;120:369.e1-7.
- [11] Fowler VG Jr, Miro JM, Hoen B, Cabell CH, Abrutyn E, Rubinstein E, Corey GR, Spelman D, Bradley SF, Barsic B, Pappas PA, Anstrom KJ, Wray D, Fortes CQ, Anguera I, Athan E, Jones P, van der Meer JT, Elliott TS, Levine DP, Bayer AS. Staphylococcus aureus endocarditis: a consequence of medical progress. *JAMA* 2005;293:3012-21.
- [12] Cabell CH, Abrutyn E, Fowler VG Jr, Hoen B, Miro JM, Corey GR, Olaison L, Pappas P, Anstrom KJ, Stafford JA, Eykyn S, Habib G, Mestres CA, Wang A. Use of surgery in patients with native valve infective endocarditis: results from the International Collaboration on Endocarditis Merged Database. *Am Heart J* 2005;150:1092-8.
- [13] Gabbieri, Dohmen PM, Linneweber J, Grubitzsch H, von Heymann C, Neumann K, Halle E, Konertz WF. Early outcome after surgery for active native and prosthetic aortic valve endocarditis. *J Heart Valve Dis* 2008;17:508-25.
- [14] Corral I, Martín-Dávila P, Fortún J, Navas E, Centella T, Moya JL, Cobo J, Quereda C, Pintado V, Moreno S. Trends in neurological complications of endocarditis. *J Neurol.* 2007;254:1253-9.
- [15] Archer NK, Mazaitis MJ, Costerton JW, Leid JG, Powers ME, Shirtliff ME. Staphylococcus aureus biofilms: properties, regulation, and roles in human disease. *Virulence.* 2011;2:445-59.
- [16] Watkin R, Sandoe J. British Society of Antimicrobial Chemotherapy (BSAC) guidelines for the diagnosis and treatment of endocarditis: what the cardiologist needs to know. *Heart.* 2012;98:757-9.
- [17] Habib G, Hoen B, Tornos P, et al. Guidelines on the prevention, diagnosis, and treatment of infective endocarditis (new version 2009): the Task Force on the Prevention, Diagnosis, and Treatment of Infective Endocarditis of the European Society of Cardiology (ESC). Endorsed by the European Society of Clinical Microbiology and Infectious Diseases (ESCMID) and the International Society of Chemotherapy (ISC) for Infection and Cancer. *Eur Heart J* 2009;30:2369-413.
- [18] Costerton JW, Geesey GG, Cheng GK. How bacteria stick. *1978 Sci Am* 238:86-95.
- [19] Hall-Stoodley L, Stoodley P, Kathju S, et al. Towards diagnostic guidelines for biofilm-associated infections. *FEMS Immunol Med Microbiol* 2012;65:127-145.
- [20] Boles BR, Horswill AR. Staphylococcal biofilm disassembly. *Trends Microbiol.* 2011;19:449-55.
- [21] O'Neill E, Pozzi C, Houston P, Smyth D, Humphreys H, Robinson DA, O'Gara JP. Association between methicillin susceptibility and biofilm regulation in Staphylococcus aureus isolates from device-related infections. *J Clin Microbiol.* 2007;45:1379-88.
- [22] Jefferson KK, Goldmann DA, Pier GB. Use of confocal microscopy to analyze the rate of vancomycin penetration through Staphylococcus aureus biofilms. *Antimicrob Agents Chemother.* 2005;49:2467-73.
- [23] Rani SA, Pitts B, Beyenal H, Veluchamy RA, Lewandowski Z, Davison WM, Buckingham-Meyer K, Stewart PS. Spatial patterns of DNA replication, protein synthesis, and oxygen concentration within bacterial biofilms reveal diverse physiological states. *J Bacteriol* 2007;189:4223-33.

- [24] Bagge N, Schuster M, Hentzer M, Ciofu O, Givskov M, Greenberg EP, Høiby N. *Pseudomonas aeruginosa* biofilms exposed to imipenem exhibit changes in global gene expression and beta-lactamase and alginate production. *Antimicrob Agents Chemother.* 2004;48:1175-87.
- [25] Oliver A, Cantón R, Campo P, Baquero F, Blázquez J. High frequency of hypermutable *Pseudomonas aeruginosa* in cystic fibrosis lung infection. *Science* 2000;288:1251-4.
- [26] Lechner S, Lewis K, Bertram R. *Staphylococcus aureus* persists tolerant to bactericidal antibiotics *J Mol Microbiol Biotechnol* 2012;22:235-44.
- [27] Bagge N, Ciofu O, Hentzer M, Campbell JI, Givskov M, Høiby N. Constitutive high expression of chromosomal beta-lactamase in *Pseudomonas aeruginosa* caused by a new insertion sequence (IS1669) located in ampD. *Antimicrob Agents Chemother.* 2002;46:3406-11.
- [28] Xu KD, McFeters GA, Stewart PS. Biofilm resistance to antimicrobial agents. *Microbiology* 2000;146:547-9.
- [29] De Beer D, Srinivasan R, Stewart PS. Direct measurement of chlorine penetration into biofilms during disinfection. *Appl Environ Microbiol* 1994;60:4339-44.
- [30] Singh R, Ray P, Das A, Sharma M. Penetration of antibiotics through *Staphylococcus aureus* and *Staphylococcus epidermidis* biofilms. *J Antimicrob Chemother* 2010;65:1955-8.
- [31] Bodí M, Ardanuy C, Olona M, Castander D, Diaz E, Rello J; Department of Critical Care, Hospital Univeristari Joan XXIII, Tarragona, Spain. Therapy of ventilator-associated pneumonia: the Tarragona strategy. *Clin Microbiol Infect.* 2001;7:32-3.
- [32] Schramm GE, Johnson JA, Doherty JA, Micek ST, Kollef MH. Methicillin-resistant *Staphylococcus aureus* sterile-site infection: The importance of appropriate initial antimicrobial treatment. *Crit Care Med.* 2006;34:2069-74.
- [33] Cheong HS, Kang CI, Wi YM, Ko KS, Chung DR, Lee NY, Song JH, Peck KR. Inappropriate initial antimicrobial therapy as a risk factor for mortality in patients with community-onset *Pseudomonas aeruginosa* bacteraemia. *Eur J Clin Microbiol Infect Dis.* 2008;27:1219-25.
- [34] Finberg RW, Moellering RC, Tally FP, Craig WA, Pankey GA, Dellinger EP, West MA, Joshi M, Linden PK, Rolston KV, Rotschafer JC, Rybak MJ. The importance of bactericidal drugs: future directions in infectious disease. *Clin Infect Dis.* 2004;39:1314-20.
- [35] Pankey GA, Sabath LD. Clinical relevance of bacteriostatic versus bactericidal mechanisms of action in the treatment of Gram-positive bacterial infections. *Clin Infect Dis.* 2004;38:864-70.
- [36] Cosgrove SE, Fowler VG Jr. Management of methicillin-resistant *Staphylococcus aureus* bacteremia. *Clin Infect Dis.* 2008;46:S386-93.
- [37] Soriano A, Marco F, Martínez JA, Pisos E, Almela M, Dimova VP, Alamo D, Ortega M, Lopez J, Mensa J. Influence of vancomycin minimum inhibitory concentration on the treatment of methicillin-resistant *Staphylococcus aureus* bacteremia. *Clin Infect Dis.* 2008;46:193-200.
- [38] Lodise TP, Graves J, Evans A, Graffunder E, Helmecke M, Lomaestro BM, Stellrecht K. Relationship between vancomycin MIC and failure among patients with methicillin-resistant *Staphylococcus aureus* bacteremia treated with vancomycin. *Antimicrob Agents Chemother.* 2008;52:3315-20.
- [39] Howden BP, Peleg AY, Stinear TP. The evolution of vancomycin intermediate *Staphylococcus aureus* (VISA) and heterogenous-VISA. *Infect Genet Evol.* 2013 pii: S1567-1348(13)00136-6.
- [40] Werth B, Steed M, Kaatz GW, Rybak MJ. Evaluation of Ceftaroline (CPT) Activity Against Heteroresistant Vancomycin Intermediate *Staphylococcus aureus* (hVISA) and VISA Methicillin-Resistant *S. aureus* (MRSA) Strains in an In Vitro Pharmacokinetic/Pharmacodynamic (PK/PD) Model: Exploring the "Seesaw Effect". *Antimicrob Agents Chemother.* 2013 Apr 1. Epub
- [41] Rolston KV, Nguyen H, Amos G, Elting L, Fainstein V, Bodey GP. A randomized double-blind trial of vancomycin versus teicoplanin for the treatment of gram-positive bacteremia in patients with cancer. *J Infect Dis.* 1994;169:350-5.
- [42] Huang JH, Hsu RB. Treatment of infective endocarditis caused by methicillin-resistant *Staphylococcus aureus*: teicoplanin versus vancomycin in a retrospective study. *Scand J Infect Dis.* 2008;40:462-7.
- [43] Wilcox MH, Tack KJ, Bouza E, Herr DL, Ruf BR, Ijzerman MM, Croos-Dabrera RV, Kunkel MJ, Knirsch C. Complicated skin and skin-structure infections and catheter-related bloodstream infections: noninferiority of linezolid in a phase 3 study. *Clin Infect Dis.* 2009 48:203-12.
- [44] Smith K, Gemmell CG, Lang S. Telavancin shows superior activity to vancomycin with multidrug-resistant *Staphylococcus aureus* in a range of in vitro biofilm models. *Eur J Clin Microbiol Infect Dis.* 2013 Apr 29. Epub.
- [45] Head SJ, Mokhles MM, Osnabrugge RL, Bogers AJ, Kappetein AP. Surgery in current therapy for infective endocarditis. *Vasc Health Risk Manag.* 2011;7:255-63.
- [46] San Román JA, López J, Revilla A, Vilacosta I, Tornos P, Almirante B, Mota P, Villacorta E, Sevilla T, Gómez I, Del Carmen Manzano M, Fulquet E, Rodríguez E, Igual A. Rationale, design, and methods for the early surgery in infective endocarditis study (ENDOVAL 1): a multicenter, prospective, randomized trial comparing the state-of-the-art therapeutic strategy versus early surgery strategy in infective endocarditis. *Am Heart J.* 2008;156:431-6.