

Rhizospheric plant-microbe interactions that enhance the remediation of contaminated soils

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The soil, as any other ecosystem, is an important habitat to thousands of organisms associated like a wide variety of fungi, actinobacteria, algae, protozoa and different types of bacteria that vary in physiology. These microorganisms can occur in association to clay particles or organic matter and in the rhizosphere of plants essential to their metabolism in synergism with plant roots; this term can be defined as the three units interacting: the plant, the soil and the microorganisms.

This manuscript reviews some of the important beneficial plant-microbe interactions that promote plant health and development, particularly those relationships between plants and plant growth-promoting rhizobacteria, plant endophytic bacteria and mycorrhizal fungi; in a way that these microorganisms increase the availability of contaminants and help plants to the extraction and removal of inorganic and organic compounds.

Keywords phytoremediation; pollutants; rhizosphere; plant growth-promoting rhizobacteria; endophytes; mycorrhizal fungi

1. Rhizosphere: a particularly microhabitat for microorganisms

The soil, as any other ecosystem, is an important habitat to thousands of organisms associate, among the microscopic ones, there is a wide variety of fungi, actinobacteria, algae, protozoa and bacteria [1]. The microorganisms can occur in association to clay particles or organic matter, in the rhizosphere of plants and in small colonies in the pores among the particles [2,3].

Microbial-plant interactions were largely investigated; however, these studies aimed mainly the plant-pathogen interactions. Only ten years ago, the ecology of microbes in the rhizosphere was focused to many kinds of decontamination processes.

The rhizosphere defined by Hiltner as the volume of soil that is influenced by the roots of plants [4,5]. and according to Lynch [6], this term can be defined as the three units interacting: the plant, the soil and the microorganisms. The composition of rhizosphere structure is highly orientated by the type of plant, quantity and composition of root exudates and different root zones [7,8].

The root-associated microorganisms establish a synergism with plant roots and can help the plant to absorb nutrients improving plant performance and consequently the quality of soils [9-11].

1.1 Soil bacteria: benefic microorganisms for plants

Bacteria may interact with and affect the growth of plants in a variety of ways. Some bacteria are phytopathogenic and actively inhibit plant growth; others (plant growth-promoting bacteria) can facilitate the growth of plants using a wide range of different mechanisms; and there are a large number of soil bacteria that do not appear to affect the growth of plants one way or the other, although this may vary as a function of a range of different soil conditions [12].

With the discovery of a number of soil microorganisms that are capable of degrading xenobiotic chemicals including herbicides, pesticides, solvents, and other organic compounds; microbial degradation might provide a reasonable and effective means of disposing of toxic chemical wastes.

Due to the sensitivity and the sequestration ability of the microbial communities to heavy metals, microbes have been used for bioremediation [13-15]; although microbial communities in metal-polluted bulk soils have been studied, there is little information on the composition of microbial community in the plant rhizosphere growing in soils highly polluted with heavy metals [16].

2. Phytoremediation a especially kind of bioremediation

Bioremediation is a technique that uses living organisms to manage or remediate polluted soils, is an emerging technology and defined as the elimination, attenuation or transformation of polluting or contaminating substances by the use of biological processes into their less toxic forms [18,19], can be applied *in situ* or *ex situ*, depending on the site that they will be applied.

Soils in the same way as aquatic environments are the target of thousands of contaminants that vary in composition and in concentration. These contaminants enter the system as a result of a wide range of actions such as intentional applications, inadequate residue disposal, accidental wastes and inappropriate use [20]. The pollution by inorganic compounds as nitrates, phosphates and perchlorates [21]; explosives such as hexahydro-1,3,5-trinitro-1,3,5-triazine (RDX) and octahydro-1,3,5,7-tetranitro-1,3,5,7-tetrazocine (HMX) [22]; monoaromatic hydrocarbons like benzene, toluene, ethylbenzene and xylene (known as BTEX) [23]; polycyclic aromatic hydrocarbons [24]; a range of herbicides such as diuron, linuron and chlorotoluron [25] and by heavy metals [26].

In the case of soil remediation, there are several factors that should be considered, such as soil characteristics; type and concentration of contaminants [27,28].

The possibility of using plants in environmental remediation is an emerging alternative to the restoration of contaminated sites [29-36]. However, this methodology requires an understanding of the intrinsic factors that contribute to its success.

Phytoremediation has become an attractive topic of research and development. Plant-assisted bioremediation, or phytoremediation, is commonly defined as the use of green or higher terrestrial plants for treating chemically or radioactively polluted soils.

Phytoremediation can be classified according to the method and/or nature of the contaminant [37-41] so; there are several names as follows:

- a) Phytoextraction: a removal process taking advantage of the unusual ability of some plants to (hyper-) absorbing and accumulating or translocating metals or metalloids to the shoots.
- b) Phytostabilization (and immobilisation): a containment process using plants often in combination with soil additives to assist plant installation to mechanically stabilizing the site and reducing pollutant transfer to other ecosystem compartments and the food chain; the organic or inorganic compound can be incorporated to the lignin or to soil humus.
- c) phytostimulation: the growing root promotes the development of rhizosphere microorganisms capable of degrading the contaminant, using exudates as carbon source.
- d) Phytovolatilisation/rhizovolatilisation: removal processes employing metabolic capabilities of plants and associated rhizosphere microorganisms to transform pollutants into volatile compounds that are released to the atmosphere. Some ions of elements of sub-groups II, V and VI of the periodic table like mercury, selenium and arsenic are absorbed by root, then converted into less toxic forms and released.
- e) phytodegradation: organic contaminants are degraded or mineralized by specific enzyme activity.
- f) rhizofiltration : use terrestrial plants to absorb, concentrate and/or precipitate contaminants in the aqueous system.

3. Assisted phytoremediation by microorganisms: rhizoremediation

It is interesting to apply plants combined to some microorganisms to increase the efficiency of contaminants extraction; such technique is called rhizoremediation [42]. Chaudhry et al. [43] report that the use of this beneficial association can work in a way that the microorganisms raise the availability of the compounds and the plants help in the extraction and removal of such compounds. It is positive for both sides, once the plants supply nutrients for microorganisms, which in turn, grow and multiply, increasing the capacity of degradation by plants or increasing the phytotoxicity of the contaminated soil [44]; however, little is known about this synergism between plants and microorganisms and the factors that generate the answers [19]. Some of the particular microorganisms that participated in the microbe-assisted phytoremediation, are describe in the next sections.

3.1 Plant growth-promoting bacteria and rhizoremediation

Plant growth-promoting bacteria may facilitate plant growth either indirectly or directly [45]. The ability of plant growth-promoting bacteria to act as biocontrol agents against phytopathogens and thus indirectly stimulate plant growth may result from any one of a variety of mechanisms including antibiotic production, depletion of iron from the rhizosphere, induced systemic resistance, production of fungal cell wall lysing enzymes, and competition for binding sites on the root [45-47]. Plant growth promoting bacteria can directly facilitate plant growth. They may fix atmospheric nitrogen and supply it to plants; synthesize siderophores which can sequester iron from the soil and provide it to plant cells which can take up the bacterial siderophore-iron complex and synthesize phytohormones such as auxins, cytokinins and gibberelins, which can act to enhance various stages of plant growth; solubilize minerals such as phosphorus, making them more readily available for plant growth.

3.1.1 Rhizoremediation of organic contaminants by PGPR's

Although PGPR was first used for prompting the plant growth and for the biocontrol of plant diseases, much attention has recently been paid on bioremediation with PGPR [48-50]. In contrast with inorganic compounds, microorganisms can degrade and even mineralize organic compounds in association with plants [51]. Bacteria capable of degrading certain kind of organic pollutant, such as polychlorinated biphenyls (PCBs) have been isolated from a range of sites and the pathways and encoding genes have also been well studied [52]. But most of these bacteria cannot survive in the near-starvation conditions found in soils, including the rhizosphere [17,53,54].

3.1.2 Rhizoremediation of metals facilitated by PGPR's

Scientists sometimes attempt to increase metal bioavailability through the addition of various chelating agents, a strategy that often works on a small scale in the laboratory but is much less effective in the field. A large number of plants have been tested for their ability to take up high levels of metals and then translocate those metals from roots to leaves and shoots, however, many so-called hyperaccumulating plants do not produce sufficient biomass to make this process efficient in the field [55]. The use of soil bacteria (often plant growth-promoting bacteria) as adjuncts in metal phytoremediation can significantly facilitate the growth of plants in the presence of high (and otherwise inhibitory) levels of metals [17,56].

3.3 Endophytic microorganisms: a particularly type of microorganisms associated to plants applied to rhizoremediation

Endophytic bacteria can be defined as bacteria colonizing the internal tissues of plants (an intimate niche) without causing symptoms of infection or negative effects on their host [57,58]. With the exception of seed endophytes, the primary site where endophytes gain entry into plants is via the roots. Several microscopic studies confirm this route of colonization [59,60]. Once inside the plant, endophytes either reside in specific plant tissues like the root cortex or the xylem or colonize the plant systematically by transport through the vascular system or the apoplast [61,62]. Endophytic bacteria have been isolated from a variety of healthy plant species ranging from herbaceous crop plants [58, 63-65] and different grass species [66,67] to woody tree species [68-70]. In general, Pseudomonaceae, Burkholderiaceae and Enterobacteriaceae are among the most common genera of cultivable endophytic species found [71].

Additionally to their beneficial effects on plant growth, endophytes have considerable biotechnological potential to improve the applicability and efficiency of phytoremediation [72].

Using both cultivation and cultivation-independent techniques, Idris et al. [73] investigated the endophytes and rhizobacteria with *Thlaspi goesingense*, a hyperaccumulator of Nickel. Generally, most of the endophytes were cultivation-independent and tolerated higher concentration of Ni than rhizosphere bacteria. Although this system is promising in heavy metal remediation, the mechanisms by which endophytes promote metal accumulation are not well understood yet and the application of cultivation-independent microbe is very difficult [72].

3.4 Mycorrhiza and plants: a new advance strategy named mycorrhizoremediation

Besides symbiotic bacteria, Fungi can also be used in phytoremediation technology, like arbuscular mycorrhizae, establishing symbiotic relationships with 80–90% land plants [54,74,75].

Mycorrhizae can efficiently explore the soil volume and, due to their small diameter, microsites that are not accessible for plant roots. They can further modify pollutant bioavailability in several ways, including competition with roots and other microorganisms for water and pollutant uptake, protection of roots from direct interaction with the pollutant via formation of the ectomycorrhizal sheath, and impeded pollutant transport through increased soil hydrophobicity [76]. Ectomycorrhizal associations can display considerable resistance against toxicity in soil polluted with metals [76,77] and organic compounds such as m-toluate [78], petroleum [79], or polycyclic aromatic hydrocarbons [80]. The structure of the fungal sheath and the density and surface area of the mycelium are likely to be important characteristics determining the efficiency of an ectomycorrhizal association to withstand metal toxicity and to protect the host plant from pollutant contact [81].

In addition to their protective role, mycorrhizae may contribute to the resistance of plant–microbial associations through enhanced degradation of organic pollutants in the mycorrhizosphere [76], thus lowering the bioavailable concentration of the pollutant in soil.

In some cases, arbuscular mycorrhizal fungi have been shown to increase uptake of metals [82-84] and arsenic [85,86] in plants but other studies showed no effect or decreased concentrations in plant tissues [87,88]. The contrasting results are difficult to evaluate and may be partly due to different experimental settings, e.g. greenhouse [85,86] versus field studies [87,88] as in the case of arsenic uptake in *Pteris vittata* inoculated with arbuscular mycorrhizal fungi [89].

4. Importance of genetically-engineered rhizobacteria to rhizoremediation

Many bacteria in the rhizosphere show only limited ability in degrading organic pollutants. With the development of molecular biology, the genetically-engineered rhizobacteria with the contaminant-degrading gene are constructed to conduct the bioremediation in rhizosphere [56].

Examples about the molecular mechanisms involved in the degradation for some pollutants such as trichloroethylene (TCE) and PCBs has been studied [52,54].

For heavy metals, Sriprang et al. [90] introduced *Arabidopsis thaliana* gene for phytochelatin synthase (PCS; PCSAt) into *Mesorhizobium huakuii* subsp. *rengei* strain B3 and then established the symbiosis between *M. huakuii* subsp. *rengei* strain B3 and *Astragalus sinicus*. The gene was expressed to produce phytochelatin and accumulate Cd²⁺, under the control of bacteroid-specific promoter, the *nifH* gene [91].

To select a suitable strain for gene recombination and inoculation into the rhizosphere, there are two criteria that has been recommended: first, the strain should be stable after cloning and the target gene should have a high expression, second, the strain should be tolerant or insensitive to the contaminant; and third, some strains can survive only in several specific plant rhizosphere [92].

5. References

- [1] Pelczar M, Reid R, Chan ECS. Microbiologia, vol. 2. São Paulo: McGraw-Hill do Brasil; 1981.
- [2] Alexander M. Introduction to soil microbiology. 2nd edition. New York: John Wiley and Sons; 1977.
- [3] Kuske CR, Ticknor LO, Miller ME, Dunbar JM, Davis JA, Barns SM. Comparison of soil bacterial communities in rhizospheres of three plant species and in the interspaces in an arid grassland. *Appl Environ Microbiol.* 2002; 68:1854–63.
- [4] Hiltner L. Über neuere Erfahrungen und Probleme auf dem Gebiet der Bodenbakteriologie und unter Besonderer Berücksichtigung der Gründung und Brache. *Arb Deutsch Landwirt Ges.* 1904; 98:59–78.
- [5] Vargas JP, Esquivel GG, García FE. Papel ecológico de la flora rizosférica en fitorremediación. *Av Perspect.* 2002; 21:297–300.
- [6] Lynch JM. The rhizosphere. New York: John Wiley and Sons; 1990.
- [7] Marschner P, Crowley D, Yang CH. Development of specific rhizosphere bacterial communities in relation to plant species, nutrition and soil type. *Plant Soil.* 2004; 261: 199–208.
- [8] Yang CH, Crowley DE. Rhizosphere microbial community structure in relation to root location and plant iron nutritional status. *Appl Environ Microbiol.* 2000; 66:345–51.
- [9] Tinker PB. The role of microorganisms in mediating and facilitating the uptake of plant nutrients from soil. *Plant Soil.* 1984; 76:77–91.
- [10] Barea JM, Azcón R, Azcón-Aguilar C. Mycorrhizosphere interactions to improve plant fitness and soil quality. *Anton Leeuw.* 2002; 81:343–51.
- [11] Yang J, Klopper JW, Ryu C-M. Rhizosphere bacteria help plants tolerate abiotic stress. *Trends Plant Sci.* 2009;14:1–4.
- [12] Glick BR. The enhancement of plant growth by free-living bacteria. *Can J Microbiol.* 1995;41:109–17.
- [13] Hallberg KB, Johnson DB. Microbiology of a wetland ecosystem constructed to remediate mine drainage from a heavy metal mine. *Sci Total Environ.* 2005; 338:53–66.
- [14] Kao PH, Huang CC, Hseu ZY. Response of microbial activities to heavy metals in a neutral loamy soil treated with biosolid. *Chemosphere.* 2006; 64:63–70.
- [15] Umrana VV. Bioremediation of toxic heavy metals using acidothermophilic autotrophes. *Bioresour Technol.* 2006; 97:1237–42.
- [16] Dell'Amico E, Cavalca L, Andreoni V. Analysis of rhizobacterial communities in perennial Gramineae from polluted water meadow soil, and screening of metal-resistant, potentially plant growth-promoting bacteria. *FEMS Microbiol Ecol.* 2005; 52:153–62.
- [17] Zhuang X, Chen J, Shim H, Bai Z. New advances in plant growth-promoting rhizobacteria for bioremediation. *Environ. Int.* 2007; 33: 406–413
- [18] Vidali M. Bioremediation. An overview. *Pure Appl Chem.* 2001;73:1163–72.
- [19] Kavamura NV, E Esposito. Biotechnological strategies applied to the decontamination of soils polluted with heavy metals. *Biotechnol Adv.* 2008; 28: 61–69.
- [20] Knaebel DB, Federle TW, McAvoy DC, Vestal JR. Effect of mineral and organic soil constituents on microbial mineralization of organic compounds in a natural soil. *Appl Environ Microbiol.* 1994;60:4500–8.
- [21] Nozawa-Inoue M, Scow KM, Rolston DE. Reduction of perchlorate and nitrate by microbial communities in vadose soil. *Appl Environ Microbiol.* 2005;71:3928–34.
- [22] Kitts CL, Cunningham DP, Unkefer PJ. Isolation of three hexahydro-1, 3, 5-trinitro-1, 3, 5-triazine-degrading species of the family Enterobacteriaceae from nitramine explosive-contaminated soil. *Appl Environ Microbiol.* 1994;60:4608–11.
- [23] Rooney-Varga JN, Anderson RT, Fraga JL, Ringelberg D, Lovley DR. Microbial communities associated with anaerobic benzene degradation in a petroleum contaminated aquifer. *Appl Environ Microbiol.* 1999;65:3056–63.
- [24] Wang X, Xiaobing Y, Bartha R. Effect of bioremediation on polycyclic aromatic hydrocarbon residues in soil. *Environ Sci Technol.* 1990; 24:1086–9.
- [25] Fantroussi S, Verschuere L, Verstraete W, Top EM. Effect of phenylurea herbicides on soil microbial communities estimated by analysis of 16S rRNA gene fingerprints and community-level physiological profiles. *Appl Environ Microbiol.* 1999; 65:982–8.
- [26] Glick BR. Phytoremediation: synergistic use of plants and bacteria to clean up the environment. *Biotechnol Adv.* 2003; 21:383–93.
- [27] Boopathy R. Factors limiting bioremediation technologies. *Biores Technol.* 2000;74:63–7.

- [28] Sheoran V, Sheoran AS, Poonam P. Remediation techniques for contaminated soils. *Environ Eng Manag. J.* 2008;7:379–87.
- [29] Singh OV, Labana S, Pandey G, Budhiraja R, Jain RK. Phytoremediation: an overview of metallic ion decontamination from soil. *Appl Microbiol Biotechnol.* 2003;61: 405–12.
- [30] Paquin DG, Campbell S, Li QX. Phytoremediation in subtropical Hawaii — a review of over 100 plant species. *Remed J.* 2004;14:127–39.
- [31] Vassilev A, Schwitzguebel J-P, Thewys T, Van Der Lelie D, Vangronsveld J. The use of plants for remediation of metal-contaminated soils. *Sci World J.* 2004;4:9-34.
- [32] Shah K, Nongkynrih JM. Metal hyperaccumulation and bioremediation. *Biol Plant.* 2007;51:618–34.
- [33] Rajkumar M, Freitas H. Influence of metal resistant-plant growth-promoting bacteria on the growth of *Ricinus communis* in soil contaminated with heavy metals. *Chemosphere.* 2008;71:834–42.
- [34] Siciliano SD, Germida JJ. Mechanisms of phytoremediation: biochemical and ecological interactions between plants and bacteria. *Environ Rev.* 1998;6:65–79.
- [35] Lone MI, He Z-H, Stoffella J, Yang X. Phytoremediation of heavy metal polluted soils and water: progresses and perspectives. *J Zhejiang Univ Sci B.* 2008;9:210–20.
- [36] Padmavathamma PK, Li LY. Phytoremediation technology: hyper-accumulation metals in plants. *Water Air Soil Pollut.* 2007;184:105–26.
- [37] Eapen S, Suseelan KN, Tivarekar S, Kotwal SA, Mitra R. Potential for rhizofiltration of uranium using hairy root cultures of *Brassica juncea* and *Chenopodium amaranticolor*. *Environ Res.* 2003; 91:127–33.
- [38] January MC, Cutright TJ, Van Keulen H, Wei R. Hydroponic phytoremediation of Cd, Cr, Ni, As, and Fe: can *Helianthus annuus* hyperaccumulate multiple heavy metals? *Chemosphere.* 2008; 70:531–7.
- [39] Lasat MM. Phytoextraction of toxic metals: a review of biological mechanisms. *J Environ Qual.* 2002; 31:109–20.
- [40] Salt DE, Blaylock M, Kumar NP, Dushenkov V, Ensley BD, Chet I, et al. Phytoremediation: a novel strategy for the removal of toxic metals from the environment using plants. *Biotechnology.* 1995; 13:468–74.
- [41] Newman LA, Reynolds CM. Phytodegradation of organic compounds. *Curr Opin Biotechnol.* 2004; 15:225–30.
- [42] Jing Y, He Z, Yang X. Role of soil rhizobacteria in phytoremediation of heavy metal contaminated soils. *J Zhejiang Univ Sci B.* 2007; 8:192–207.
- [43] Chaudhry Q, Blom-Zandstra M, Gupta SK, Joner E. Utilizing the synergy between plants and rhizosphere microorganisms to enhance breakdown of organic pollutants in the environment. *Environ Sci Pollut Res.* 2005; 12:34–48.
- [44] Siciliano SD, Germida JJ. Mechanisms of phytoremediation: biochemical and ecological interactions between plants and bacteria. *Environ Rev.* 1998;6:65–79.
- [45] Glick BR, Karaturovic D, Newell P. A novel procedure for rapid isolation of plant growth promoting rhizobacteria. *Can J Microbiol.* 1995; 41:533–6.
- [46] Glick BR, Todorovic B, Czarny J, Cheng Z, Duan J, McConkey B. Promotion of plant growth by bacterial ACC deaminase. *Crit Rev Plant Sci.* 2007; 26:227–42.
- [47] Glick BR, Cheng Z, Czarny J, Duan J. Promotion of plant growth by ACC deaminase containing soil bacteria. *Eur J Plant Pathol.* 2007; 119:329–39.
- [48] Huang XD, El-Alawi Y, Penrose DM, Glick BR, Greenberg BM. A multiprocess phytoremediation system for removal of polycyclic aromatic hydrocarbons from contaminated soils. *Environ Pollut.* 2004; 130:465–76.
- [49] Huang XD, El-Alawi Y, Gurska J, Glick BR, Greenberg BM. A multi-process phytoremediation system for decontamination of persistent total petroleum hydrocarbons (TPHs) from soils. *Microchem J.* 2005; 81:139–47.
- [50] Narasimhan K, Basheer C, Bajic VB, Swarup S. Enhancement of plant-microbe interactions using a rhizosphere metabolomics-driven approach and its application in the removal of polychlorinated biphenyls. *Plant Physiol.* 2003; 132:146–53.
- [51] Saleh S, Huang XD, Greenberg BM, Glick BR. Phytoremediation of persistent organic contaminants in the environment. In: Singh A, Ward O, editors. *Soil Biology: vol. 1. Applied Bioremediation and Phytoremediation.* Berlin: Springer-Verlag; 2004. p. 115–34.
- [52] Brazil GM, Kenefick L, Callanan M, Haro A, de Lorenzo V, Dowling DN. Construction of a rhizosphere pseudomonad with potential to degrade polychlorinated biphenyls and detection of bph gene expression in the rhizosphere. *Appl Environ Microbiol.* 1995; 61:1946–52.
- [53] Normander B, Hendriksen NB, Nybroe O. Green fluorescent protein-marked *Pseudomonas fluorescens*: localization, viability, and activity in the natural barley rhizosphere. *Appl Environ Microbiol.* 1999; 65:4646–51.
- [54] Huang XD, El-Alawi Y, Penrose DM, Glick BR, Greenberg BM. Responses of three grass species to creosote during phytoremediation. *Environ Pollut.* 2004;130:453–63.
- [55] Raskin I, Ensley BD. *Phytoremediation of Toxic Metals: Using Plants to Clean Up the Environment.* New York: Wiley-Interscience; 2000.
- [56] Glick BR. Using soil bacteria to facilitate phytoremediation. *Biotechnol Adv.* 2010; 28:367–374.
- [57] Schulz B, Boyle C. What are endophytes? In: Schulz BJE, Boyle CJC, eds. *Microbial Root Endophytes.* Sieber TN. 2006.
- [58] Lodewyckx C, Mergeay M, Vangronsveld J, Clijsters H, van der Lelie D. Isolation, characterization, and identification of bacteria associated with the zinc hyperaccumulator *Thlaspi caerulescens* subsp. *calaminaria*. *Int J Phytoem.* 2002; 4:101–115.
- [59] Pan MJ, Rademan S, Kuner K, Hastings JW. Ultrastructural studies on the colonization of banana tissue and *Fusarium oxysporum* f. sp. *cubense* race 4 by the endophytic bacterium *Burkholderia cepacia*. *J Phytopathol.* 1997; 145:479–486.
- [60] Germaine K, Keogh E, Borremans B, van der Lelie D, Barac T, Oeyen L, Vangronsveld J, Porteus Moore F, Moore ERB, Campbell CD. Colonization of poplar trees by GFP expressing endophytes. *FEMS Microbiol Ecol.* 2004; 48:109–118.
- [61] Mahaffee WF, Klopper JW, Van Vuurde JW, Van der Wolf JM, Van den Brink M. Endophytic colonization of *Phaseolus vulgaris* by *Pseudomonas fluorescens* strain 89B-27 and *Enterobacter asburiae* strain JM22. In: Ryder MHR, Stevens PM, Bowen GD, eds. *Improving Plant Productivity in Rhizosphere Bacteria.* 1997.
- [62] Quadt-Hallmann A, Benhamou N, Klopper JW. Bacterial endophytes in cotton: mechanisms entering the plant. *Can J Microbiol.* 1997; 43:577–582.

- [63] Lodewyckx C, Taghavi S, Mergeay M, Vangronsveld J, Clijsters H, van der Lelie D. The effect of recombinant heavy metal resistant endophytic bacteria on heavy metal uptake by their host plant. *Int J Phytorem.* 2001; 3:173-187.
- [64] Malinowski DP, Zuo H, Belesky DP, Alloush GA. Evidence for copper binding by extracellular root exudates of tall fescue but not perennial ryegrass infected with *Neotyphodium* spp. endophytes. *Plant Soil.* 2004; 267:1-12.
- [65] Mastretta C, Taghavi S, van der Lelie D, Mengoni A, Galardi F, Gonnelli C, Barac T, Boulet J, Weyens N, Vangronsveld J. Endophytic bacteria from seeds of *Nicotiana tabacum* can reduce Cd phytotoxicity. *Int J Phytoremediat.* 2009; 11:251-267
- [66] Zinniel DK, Lambrecht P, Harris NB, Feng Z, Kuczmarski D, Higley P, Ishimaru CA, Arunakumari A, Barletta RG, Vidaver AK. Isolation and characterization of endophytic colonizing bacteria from agronomic crops and prairie plants. *Appl Environ Microbiol.* 2002; 68:2198-2208.
- [67] Dalton D, Kramer S, Azios N, Fusaro S, Cahill E, Kennedy C. Endophytic nitrogen fixation in dune grasses (*Ammophila arenaria* and *Elymus mollis*) from Oregon. *FEMS Microbiol Ecol.* 2004; 49:469-479.
- [68] Porteous Moore F, Barac T, Borremans B, Oeyen L, Vangronsveld J, van der Lelie D, Campbell CD, Moore ERB. Endophytic bacterial diversity in Poplar trees growing on a BTEX-contaminated site: the characterization of isolates with potential to enhance phytoremediation. *Sys App Micro.* 2006; 29:539-556.
- [69] Cankar K, Kraigher H, Ravnkar M, Rupnik M. Bacterial endophytes from seeds of Norway spruce (*Picea abies* L. Karts). *FEMS Microbiol Lett.* 2005; 244:341-345.
- [70] Taghavi S, Garafola C, Monchy S, Newman L, Hoffman A, Weyens N, Barac T, Vangronsveld J, van der Lelie D. Genome survey and characterization of endophytic bacteria exhibiting a beneficial effect on growth and development of poplar. *Appl Environ Microbiol.* 2009; 75:748-757.
- [71] Mastretta C, Barac T, Vangronsveld J, Newman L, Taghavi S, van der Lelie D. Endophytic bacteria and their potential application to improve the phytoremediation of contaminated environments. *Biotech Gen Eng Rev.* 2006; 23:175-207.
- [72] Weyens N, D van der Lelie, S Taghavi, J Vangronsveld. Phytoremediation: plant-endophyte partnerships take the challenge. *Curr Opin Biotechnol.* 2009; 20:1-7.
- [73] Idris R, Trifonova R, Puschenreiter M, Wenzel WW, Sessitsch A. Bacterial communities associated with flowering plants of the Ni hyperaccumulator *Thlaspi goesingense*. *Appl Environ Microbiol.* 2004; 70:2667-77.
- [74] Khan AG. Role of soil microbes in the rhizospheres of plants growing on trace metal contaminated soils in phytoremediation. *J Trace Elem Med Biol.* 2005; 18:355-64.
- [75] Khan AG. Mycorrhizoremediation - an enhanced form of phytoremediation. *J Zhejiang Univ Sci B.* 2006;7:503-14.
- [76] Meharg AA, Cairney JWG. Extomycorrhizas- extending the capabilities of rhizosphere remediation. *Soil Biol Biochem.* 2000; 32:1475-1484.
- [77] Leyval C, Turnau K, Haselwandter K. Effect of heavy metal pollution on mycorrhizal colonization and function: physiological, ecological and applied aspects. *Mycorrhiza.* 1997; 7:139-153.
- [78] Sarand I, Timonen S, Nurmiäho-Lassila E-L, Koivula T, Haahtela K, Romantschuk M. Microbial biofilms and catabolic plasmid harbouring degradative fluorescent pseudomonads in Scots pine ectomycorrhizospheres developed on petroleum contaminated soil. *FEMS Microbiol Ecol.* 1998; 27:115-126.
- [79] Sarand I, Timonen S, Koivula T, Peltola R, Haahtela K, Sen R. Tolerance and biodegradation of m-toluate by Scots pine, a mycorrhizal fungus and fluorescent pseudomonads individually and under associative conditions. *J Appl Microbiol.* 1999; 86:817-826.
- [80] Leyval C, Binet P. Effect of polycyclic aromatic hydrocarbons in soil on arbuscular mycorrhizal plants. *J Environ Qual.* 1998; 27:402-407.
- [81] Hartley J, Cairney JWG, Meharg AA. Do ectomycorrhizal fungi exhibit adaptive tolerance to potentially toxic metals in the environment? *Plant Soil.* 1997; 189:303-319.
- [82] Liao JP, Lin XG, Cao ZH, Shi YQ, Wong MH. Interactions between arbuscular mycorrhizae and heavy metals under sand culture experiment. *Chemosphere.* 2003; 50:847-853.
- [83] Whitfield L, Richards AJ, Rimmer DL. Effects of mycorrhizal colonization on *Thymus polytrichus* from heavy-metal-contaminated sites in northern England. *Mycorrhiza.* 2004; 14:47-54.
- [84] Citterio S, Prato N, Fumagalli P, Aina R, Massa N, Santagostino A. The arbuscular mycorrhizal fungus *Glomus mosseae* induces growth and metal accumulation changes in *Cannabis sativa* L. *Chemosphere.* 2005; 59:21-29.
- [85] Liu Y, Zhu YG, Chen BD, Christie P, Li XL. Influence of the arbuscular mycorrhizal fungus *Glomus mosseae* on uptake of arsenate by the As hyperaccumulator fern *Pteris vittata* L. *Mycorrhiza.* 2005; 15:187-192.
- [86] Leung HM, Ye ZH, Wong MH. Interactions of mycorrhizal fungi with *Pteris vittata* (As hyperaccumulator) in As-contaminated soils. *Environ Pollut.* 2006; 139:1-8.
- [87] Trotta A, Falaschi P, Cornara L, Minganti V, Fusconi A, Drava G. Arbuscular mycorrhizae increase the arsenic translocation factor in the As hyperaccumulating fern *Pteris vittata* L. *Chemosphere.* 2006; 65:74-81.
- [88] Wu FY, Ye ZH, Wu SC, Wong MH. Metal accumulation and arbuscular mycorrhizal status in metallicolous and nonmetallicolous populations of *Pteris vittata* L. and *Sedum alfredii* Hance. *Planta.* 2007; 226:1363-1378.
- [89] Wenzel WW. Rhizosphere processes and management in plant-assisted bioremediation (phytoremediation) of soils. *Plant Soil.* 2009; 321:385-408.
- [90] Sriprang R, Hayashi M, Ono H, Takagi M, Hirata K, Murooka Y. Enhanced accumulation of Cd²⁺ by a *Mesorhizobium* sp. transformed with a gene from *Arabidopsis thaliana* coding for phytochelatin synthase. *Appl Environ Microbiol.* 2003; 69:1791-1796.
- [91] Perret X, Freiberg C, Rosenthal A, Broughton WJ, Fellay R. High-resolution transcriptional analysis of the symbiotic plasmid of *Rhizobium* sp. NGR234. *Mol Microbiol.* 1999;32: 415-425.
- [92] Yee DC, Maynard JA, Wood TK. Rhizoremediation of trichloroethylene by a recombinant, root-colonizing *Pseudomonas fluorescens* strain expressing toluene ortho-monoxygenase constitutively. *Appl Environ Microbiol.* 1998; 64:112-118.